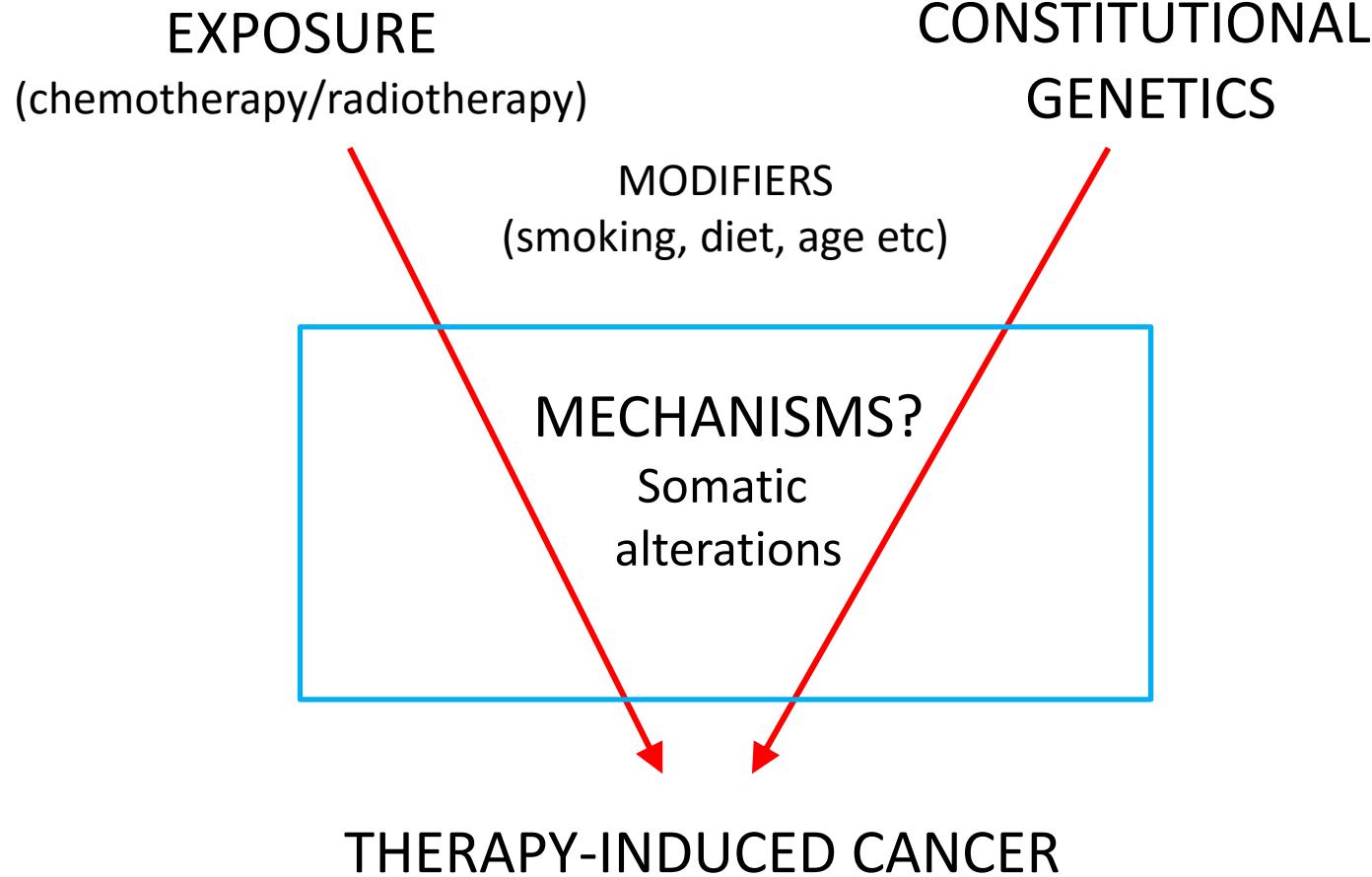
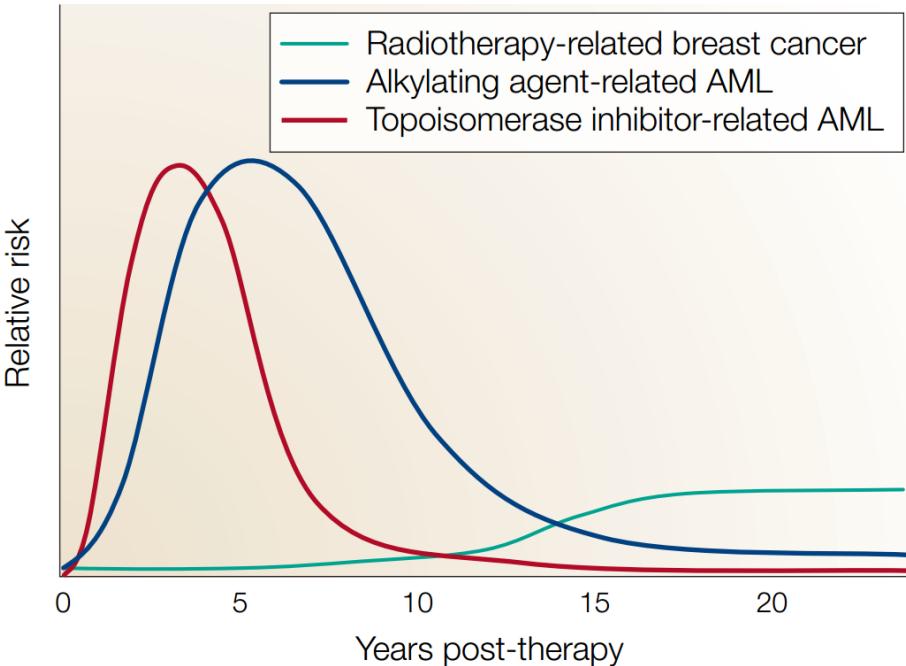


Molecular Mechanisms of Therapy-Induced Cancer

James M. Allan



Chemotherapy-induced AML



Relative risk of developing a therapy-associated cancer after Hodgkin lymphoma

ALKYLATING AGENTS

(procarbazine, dacarbazine, temozolomide, cyclophosphamide, carmustine etc)

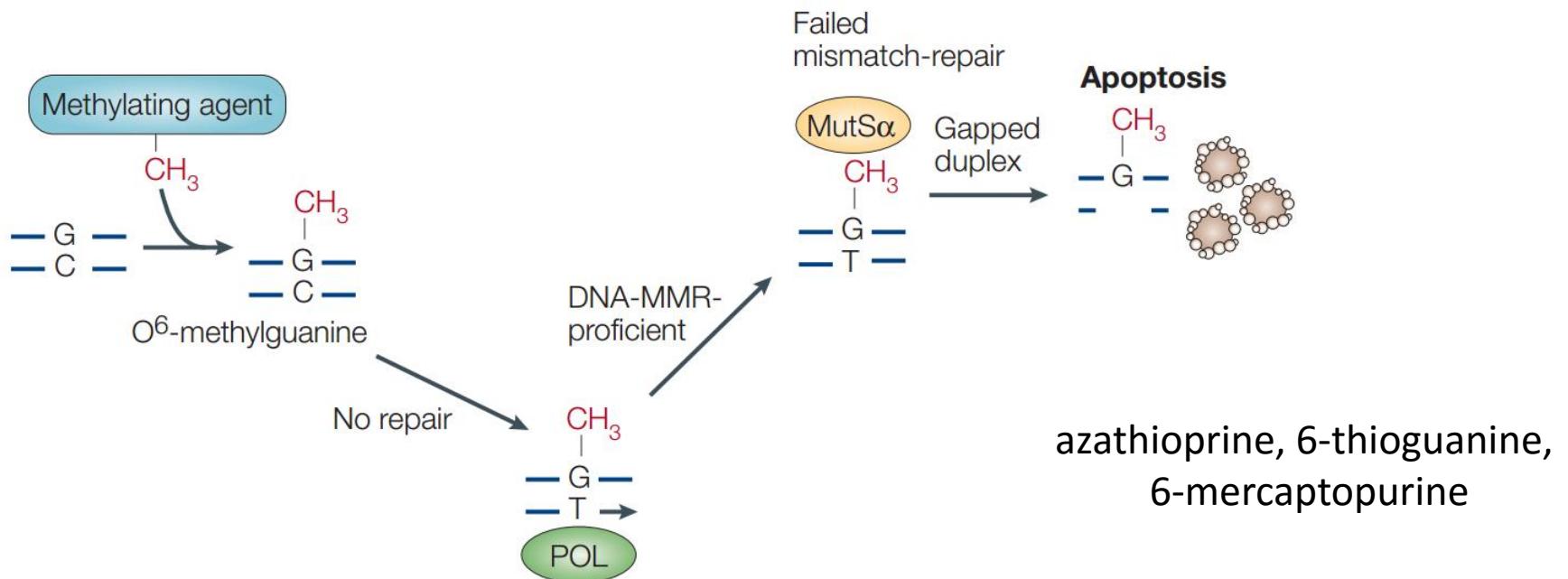
- Risk increases 1-2 yr, peaks 5-10 yr, then decreases
- Preceding myelodysplastic syndrome (4-7 hits)
- Chromosomal deletions or monosomies (Chr. 5/7)
- DNA mismatch repair defective with high mutation rate (after methylating agents)

TOPOISOMERASE INHIBITORS

(etoposide, teniposide, doxorubicin, daunorubicin etc)

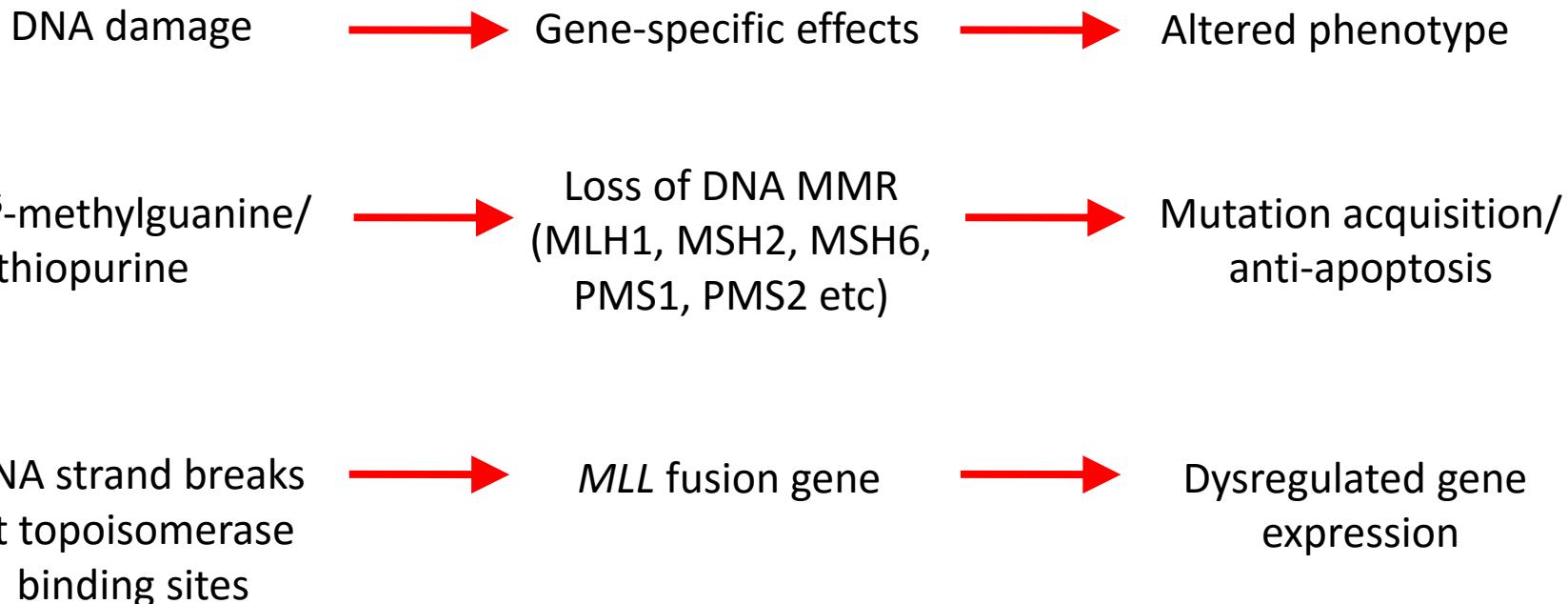
- Shorter induction period (median 2-3 yr)(1-2 hits)
- Balanced chromosomal translocations (*MLL* at 11q23)(*MLL* breaks map to topoisomerase binding sites)

Methylating agents select for DNA mismatch repair loss



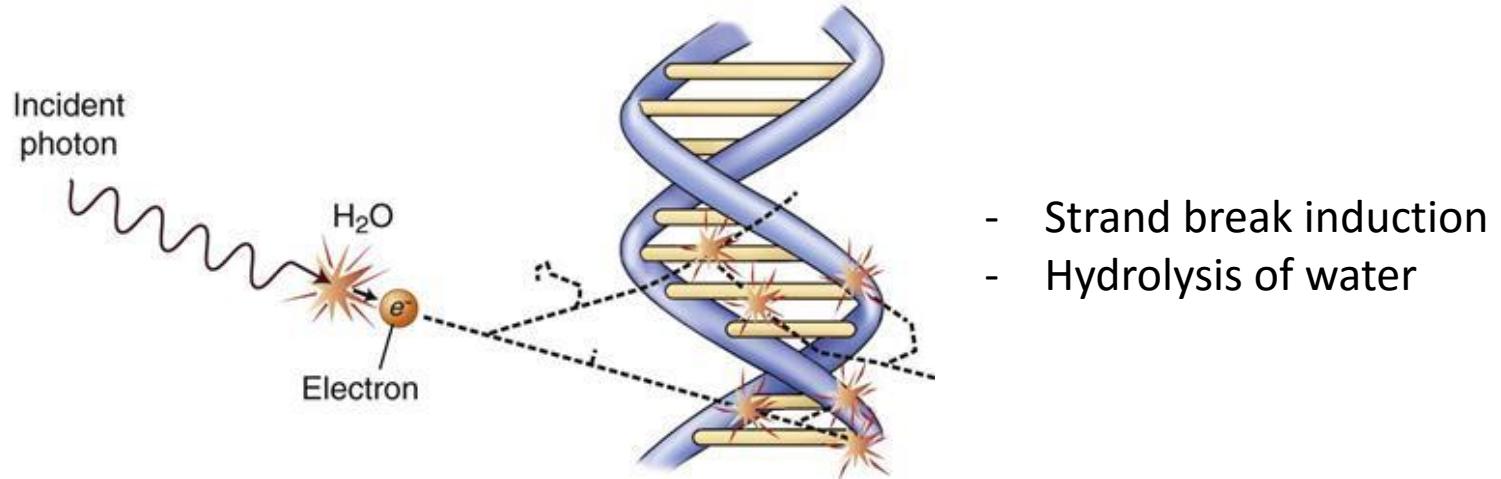
mutation
rate ↑

Molecular mechanisms of transformation



IONISING RADIATION?

Radiation induced cellular damage



- Strand break induction
- Hydrolysis of water

Radiation exposure induces DNA double-strand breaks and DNA base
– cytotoxic AND mutagenic –

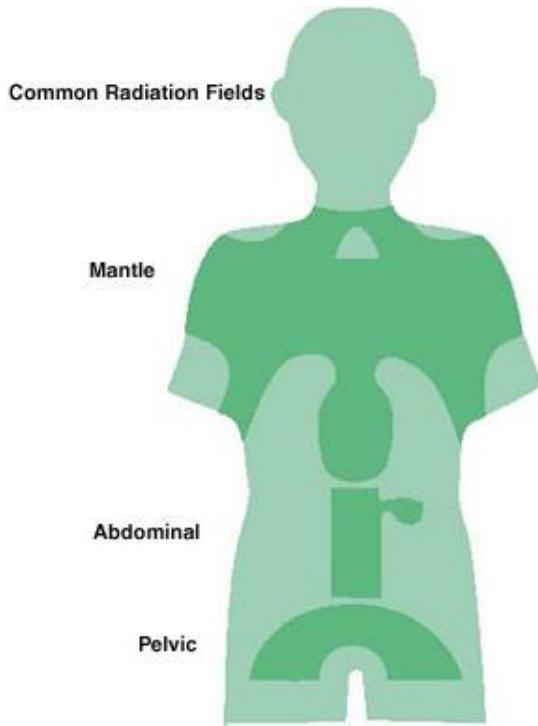
Clustered DNA damage difficult to repair and can be converted to DNA DSBs.

DNA DSBs difficult to repair with high fidelity – translocations, loss (deletion) and gain (amplification) of genetic material.

How do we identify genetic lesions induced by radiation?

MODELS !

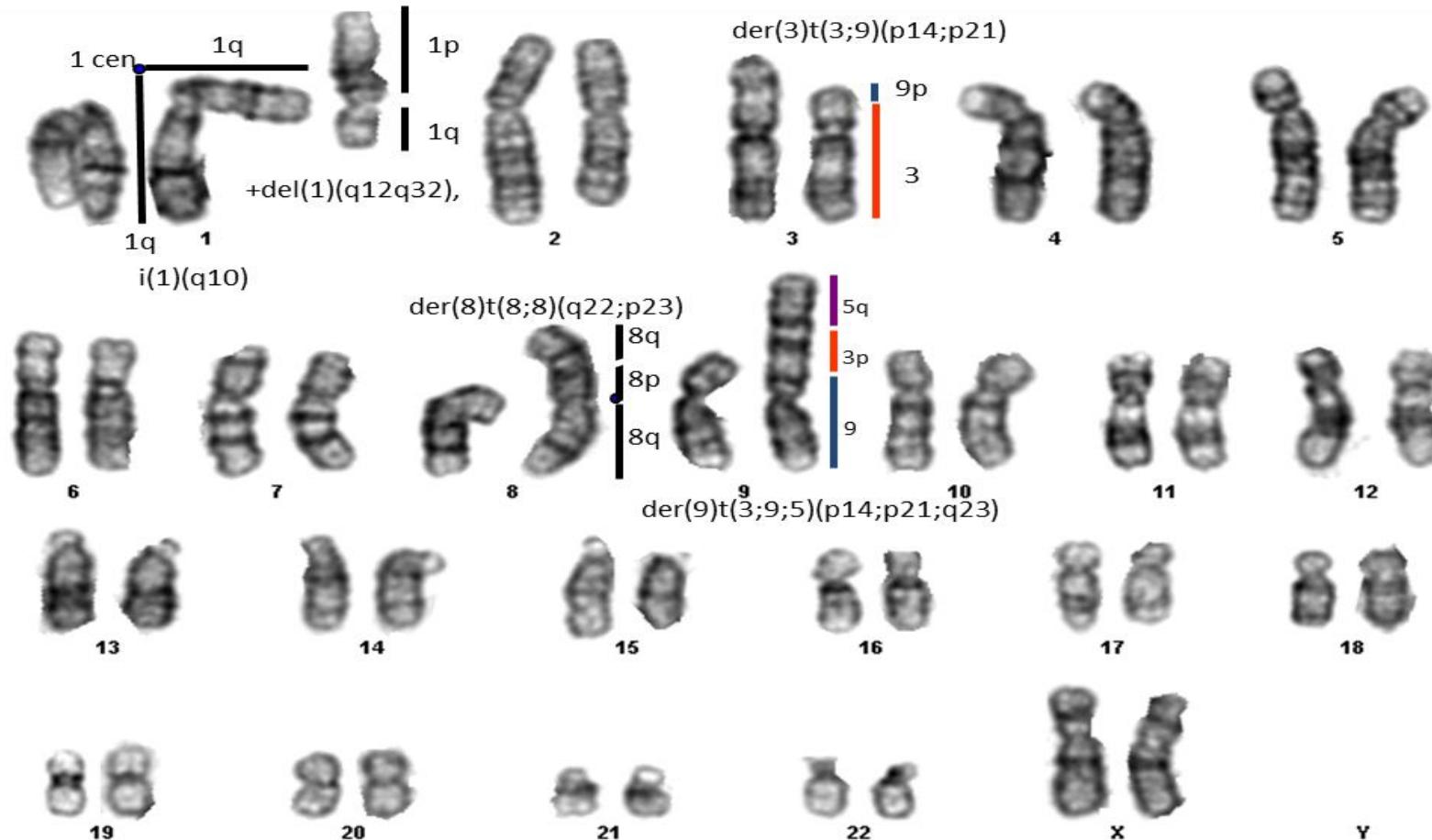
Radiogenic cancer in Hodgkin lymphoma survivors



- Risk of second cancer associated with radiation exposure
 - Positively correlated with dose
 - Inversely correlated with age
- Most common second cancers
 - **Breast**
 - Thyroid
- Contribution to risk of chemotherapy is unclear (site-specific?)
- Responsible for 20% of the mortality in HL survivors

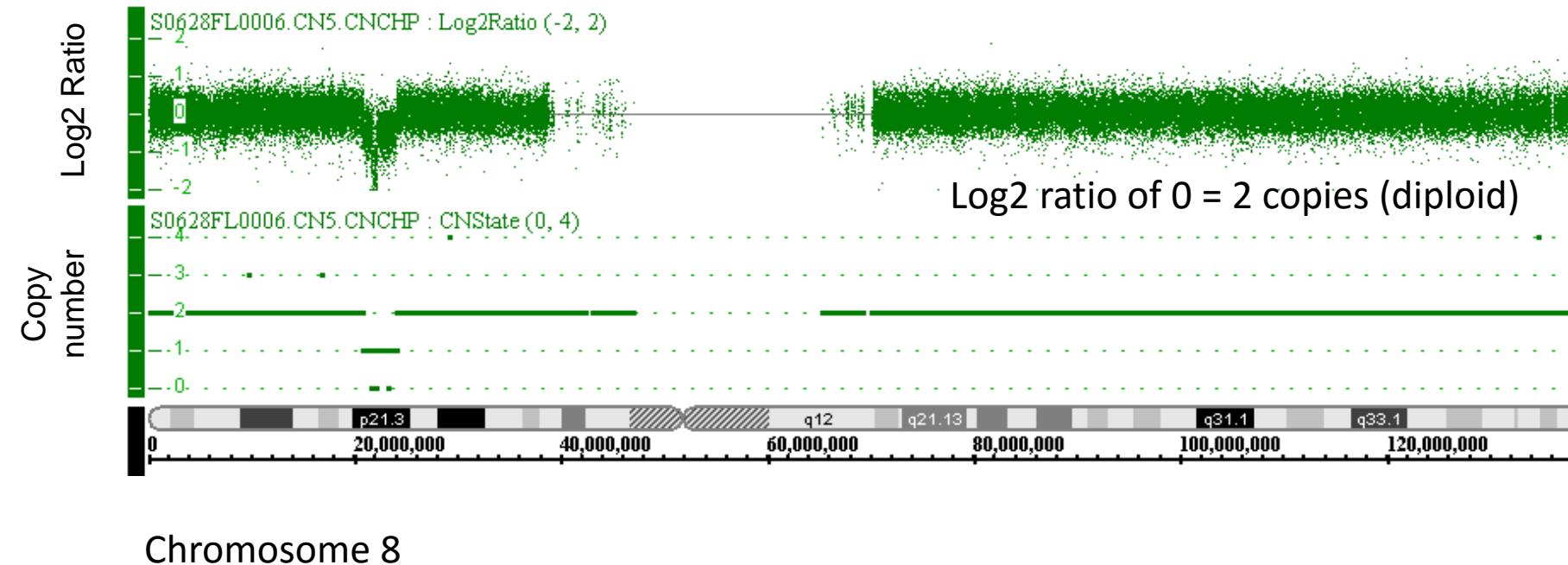
In vitro model system based on the MCF10A breast epithelial cell line

47, XX, i(1)(q10),+del(1)(q12q32), der(3)t(3;9)(p14;p21), der(8)t(8;8)(q22;p23), der(9)t(3;9;5)(p14;p21;q23).



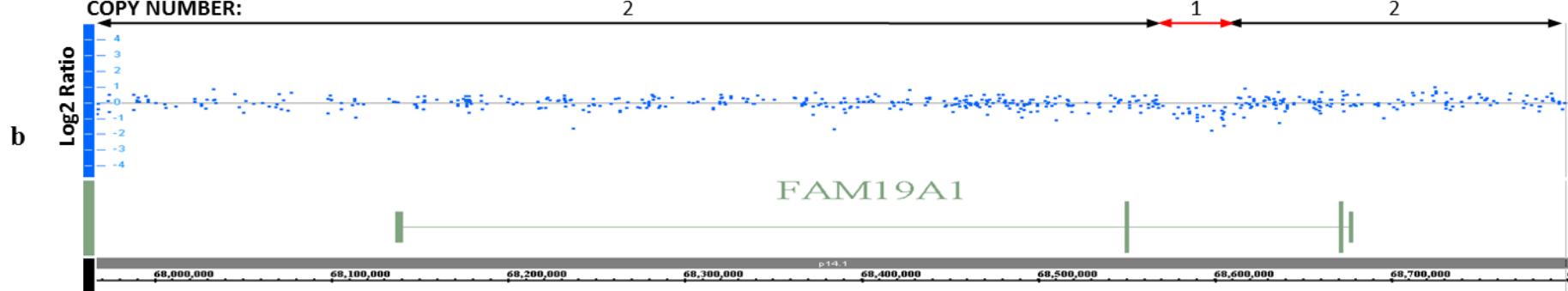
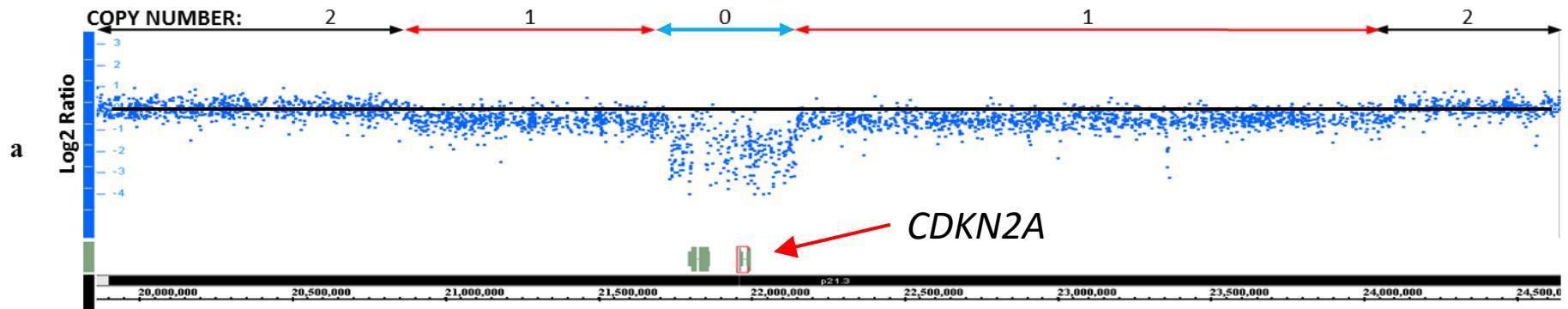
- reciprocal translocation between chromosomes 3 and 9 (deletion of *CDKN2A/2B*)
- non-reciprocal duplication of the end of the q arm of chromosome 5 to the derivative chromosome 9
- Isochromosome 1 and 8q duplication

SNP array karyotyping



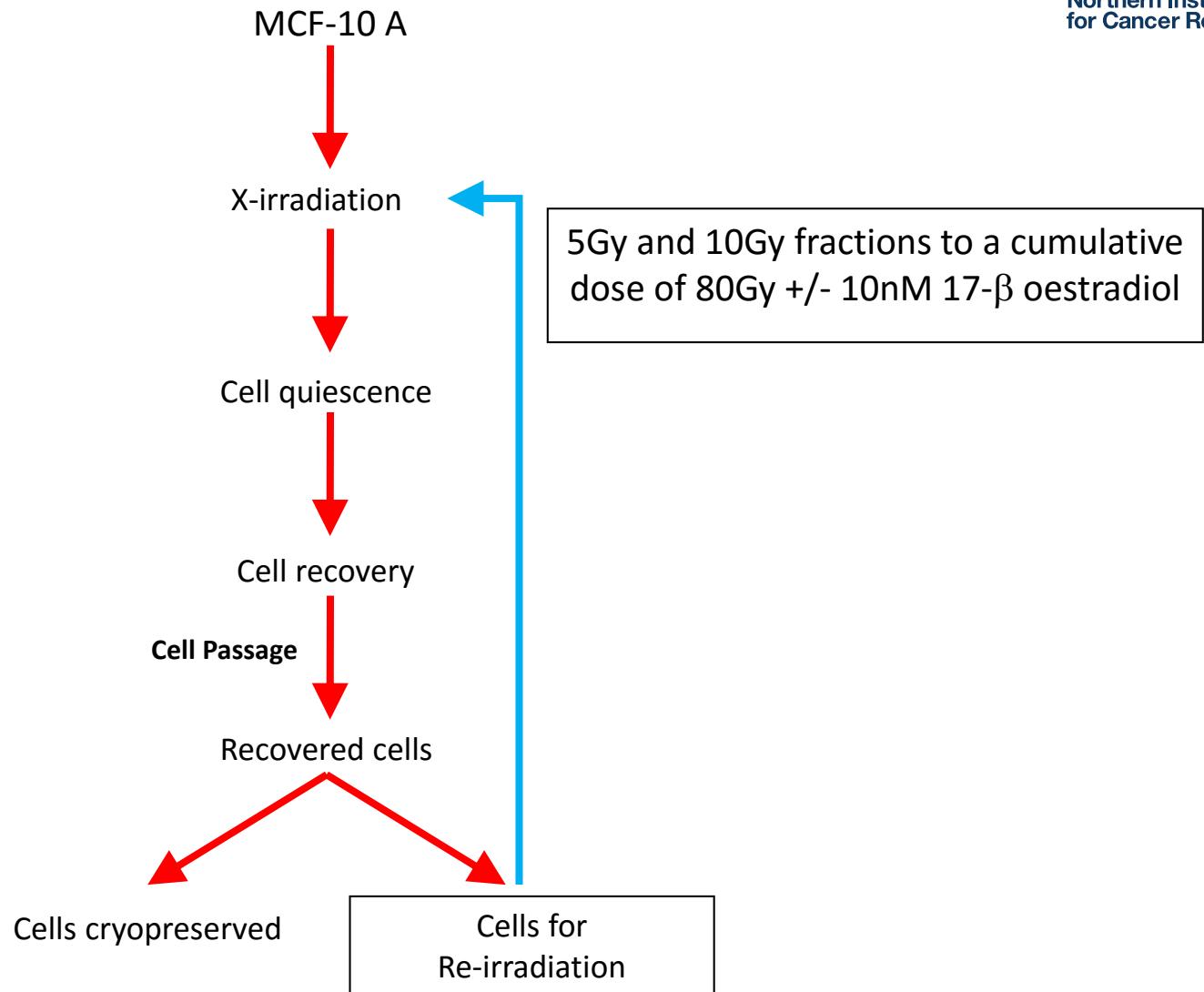
Immortality conferred by *CDKN2A* deletion

Chromosome 9 breakpoint



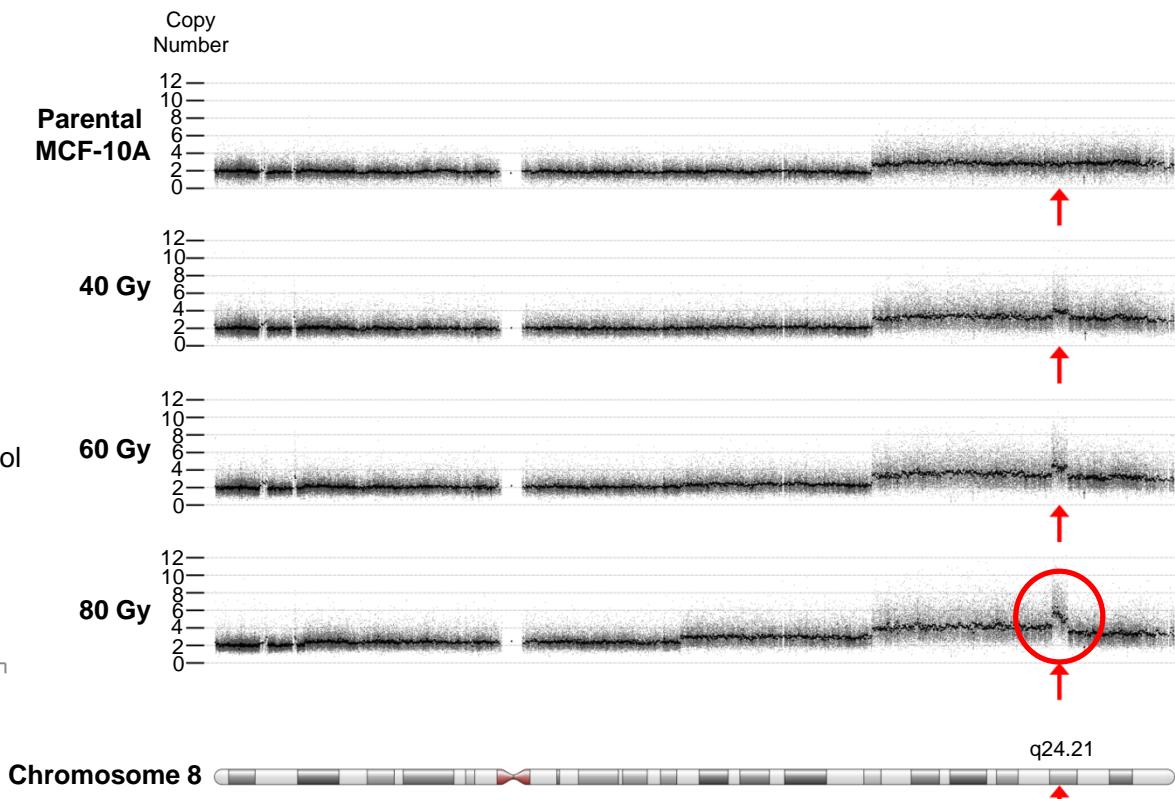
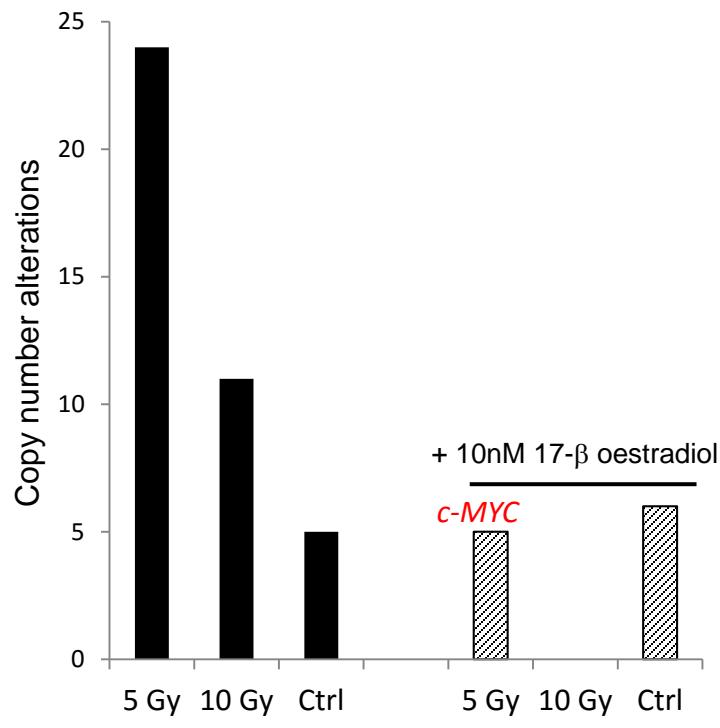
Chromosome 3 breakpoint

Irradiation protocol



Radiogenic copy number alterations

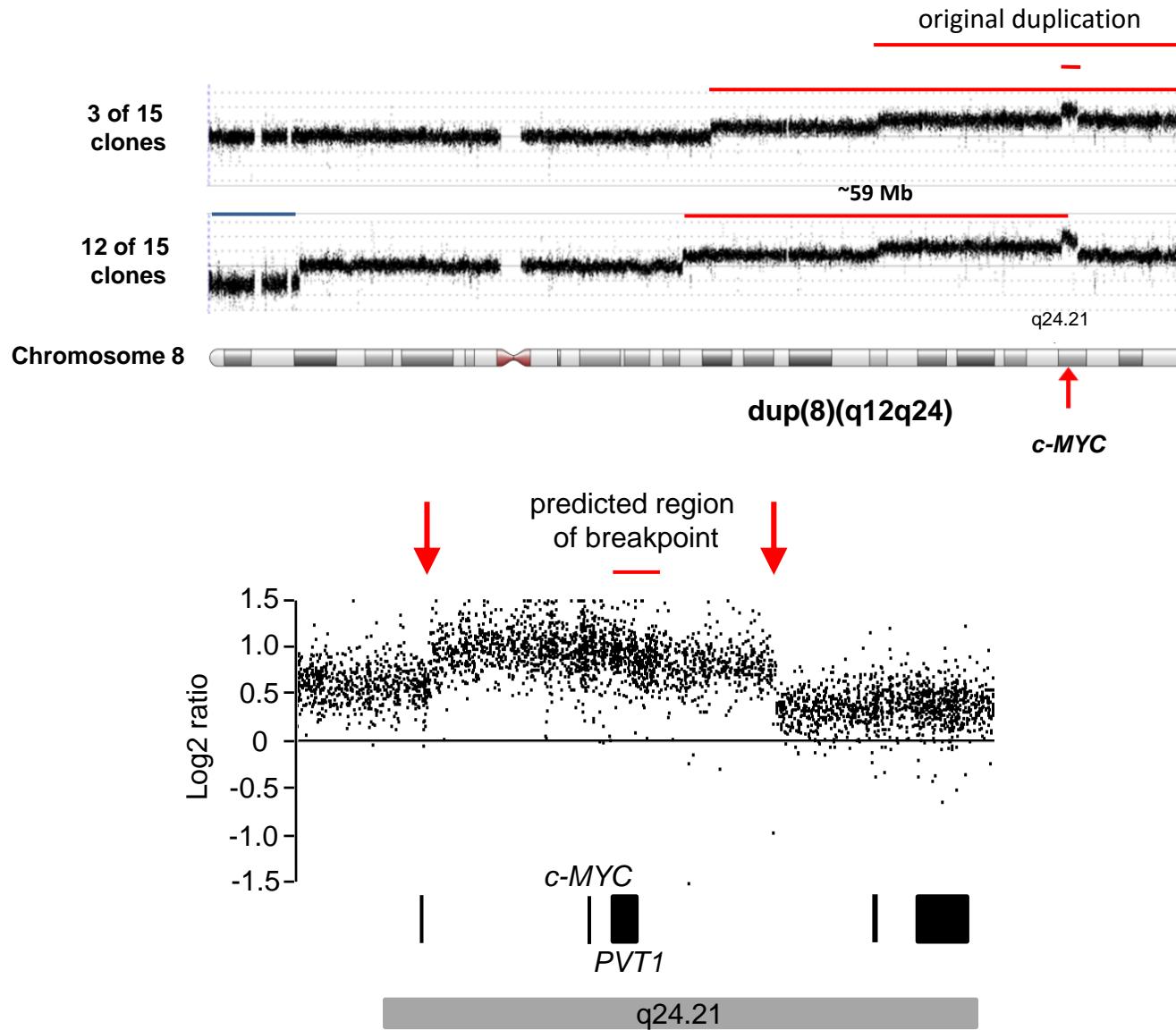
>50kb discernible



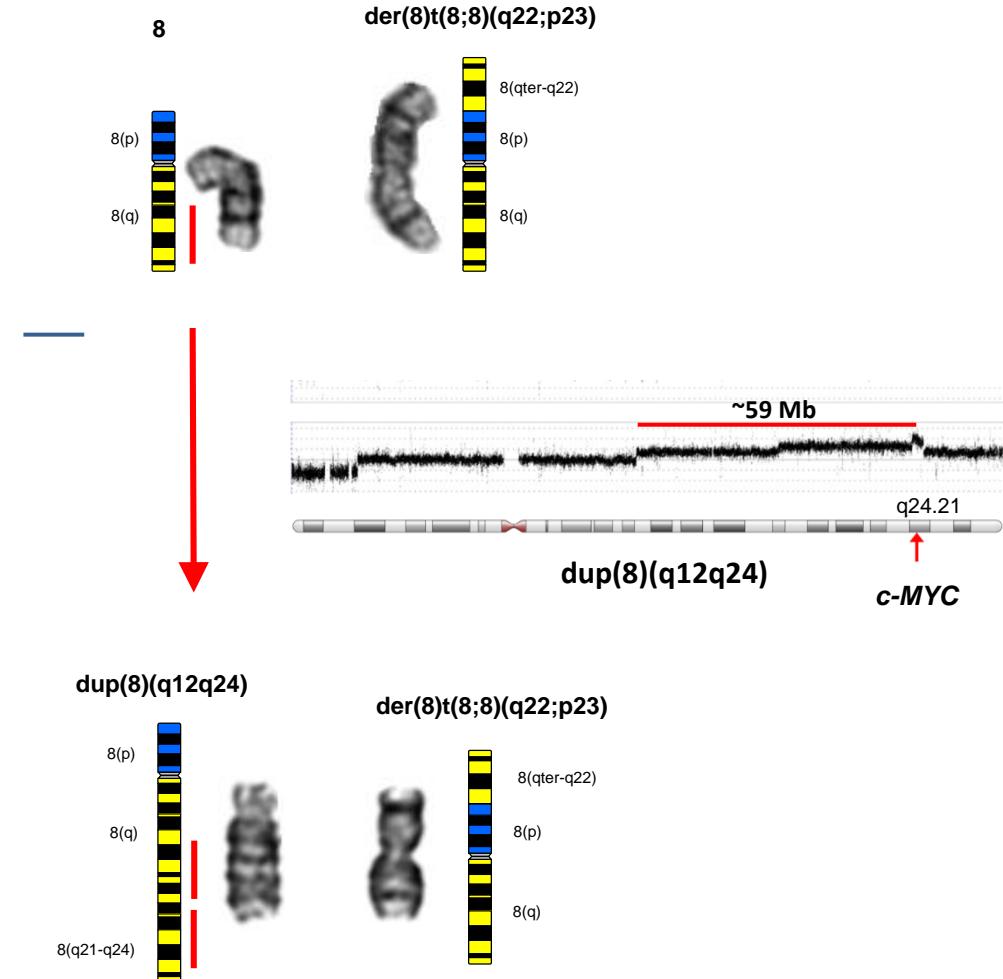
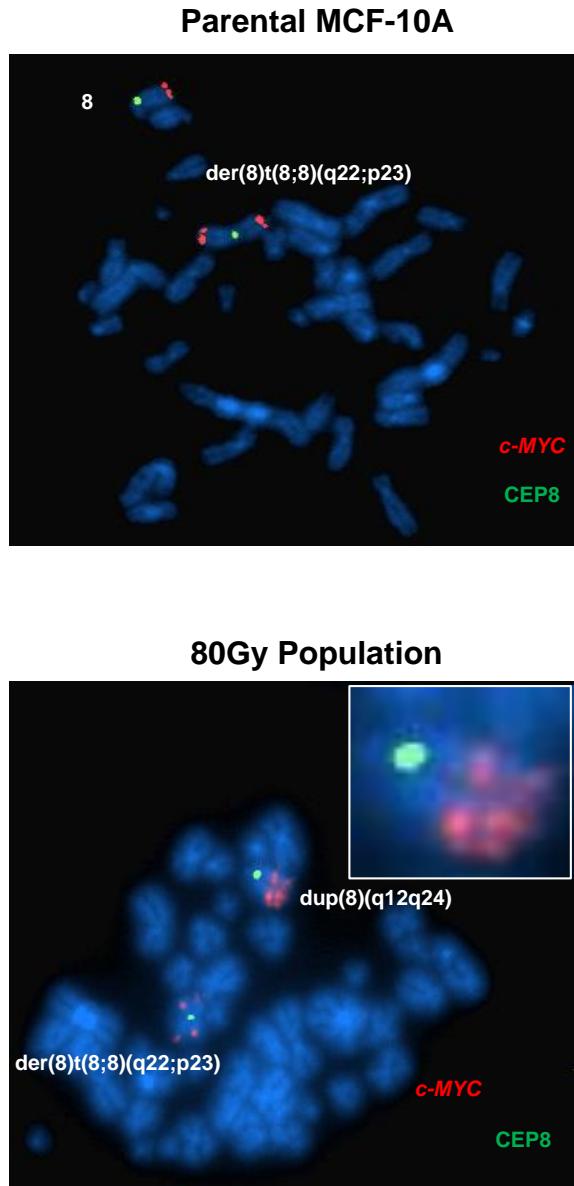
c-MYC.....

- has a pivotal function in growth control, differentiation and apoptosis.
- over-expression has potent oncogenic activity.
- dysregulation is a hallmark of many cancer types.

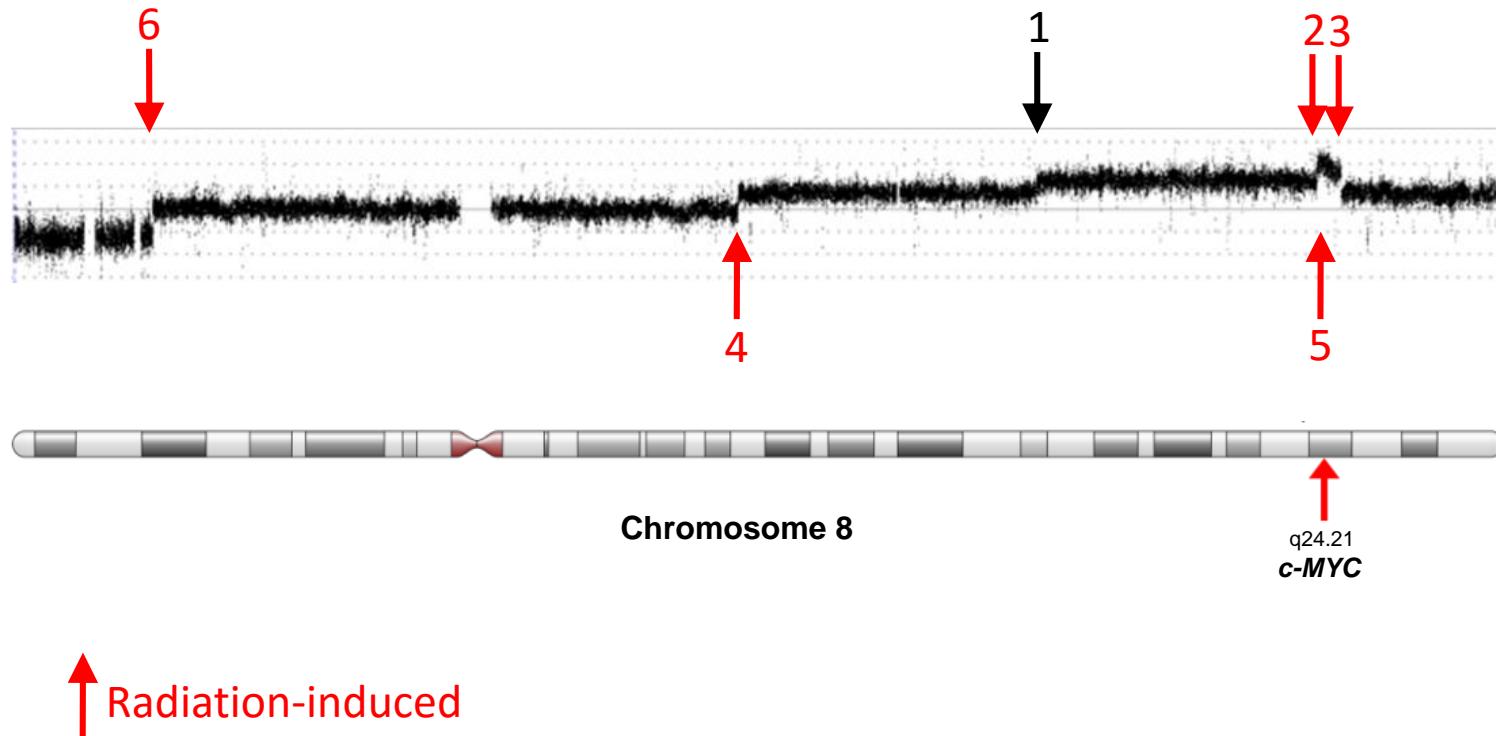
Evolution of the *c-MYC* locus in irradiated cells



Evolution of the *c-MYC* locus in irradiated cells

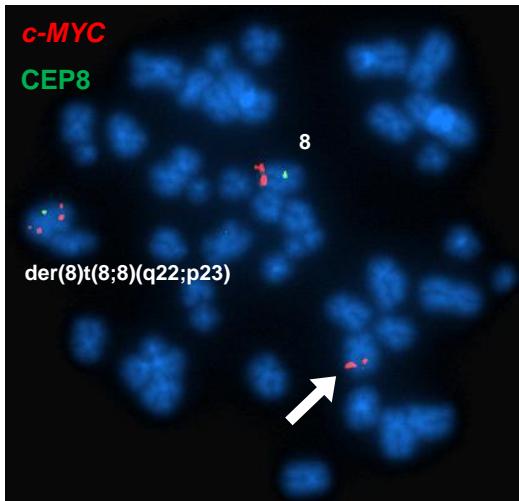


Evolution of the *c-MYC* locus in irradiated cells

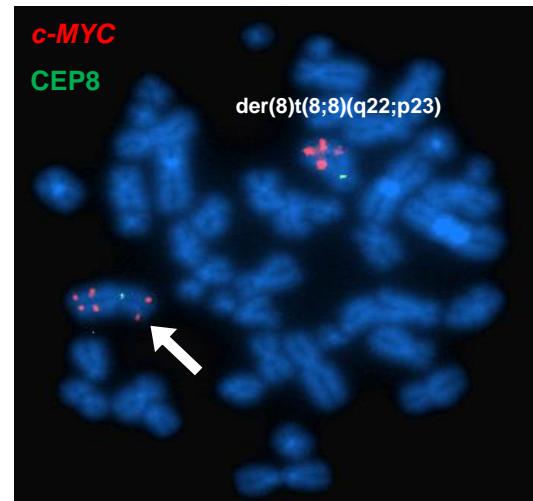


c-MYC is prone to alteration in irradiated cells

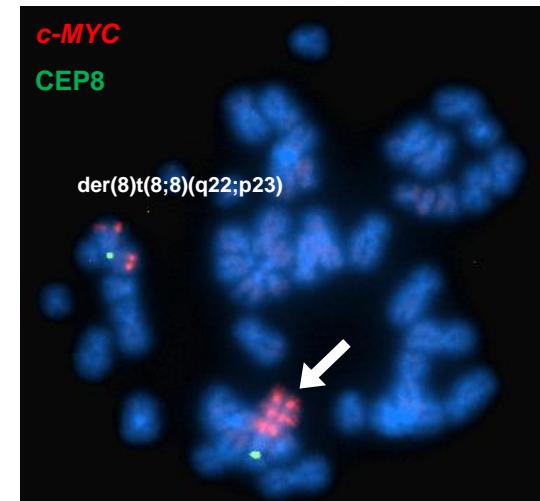
Translocation



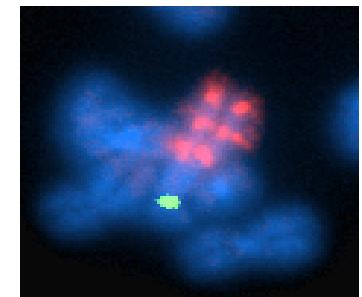
Duplication



Tripllication

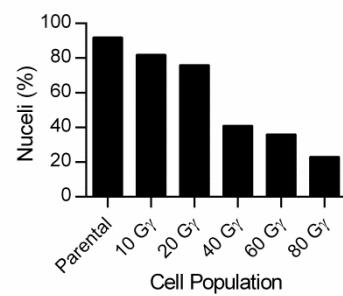
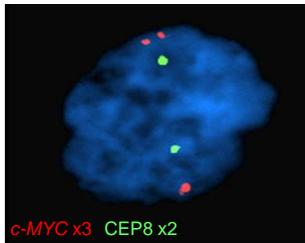


Rare events – not part of the major clone

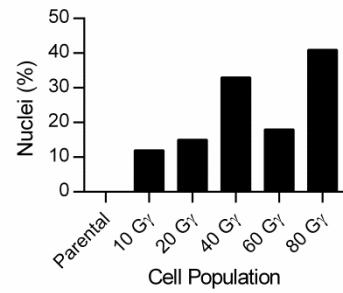
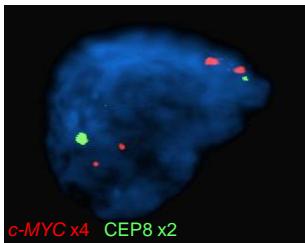


Radiation selects for *c-MYC* amplified cells

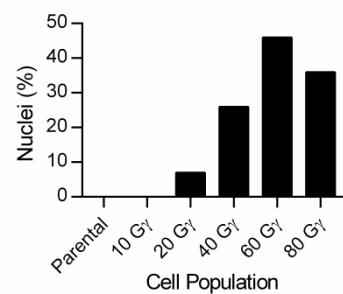
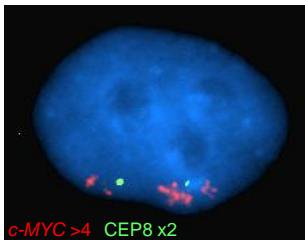
3 *c-MYC* copies (MCF-10A)



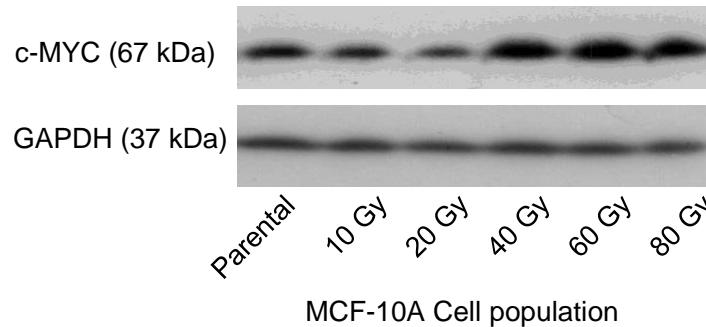
4 *c-MYC* copies (MCF-10A)



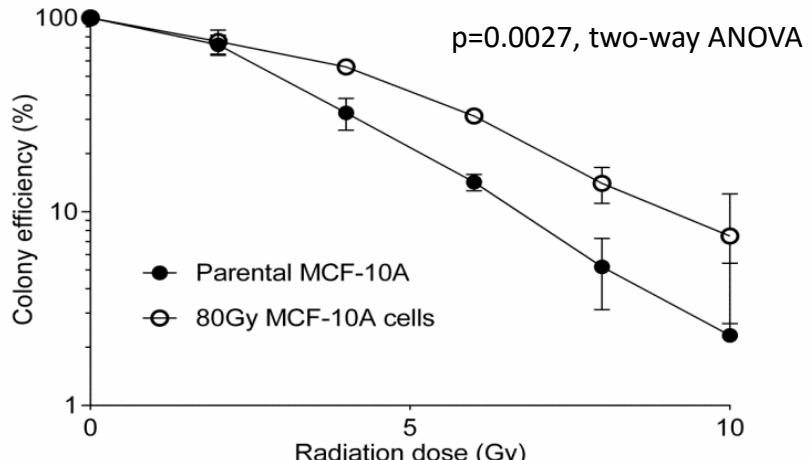
>4 *c-MYC* copies (MCF-10A)



Amplification correlated with expression

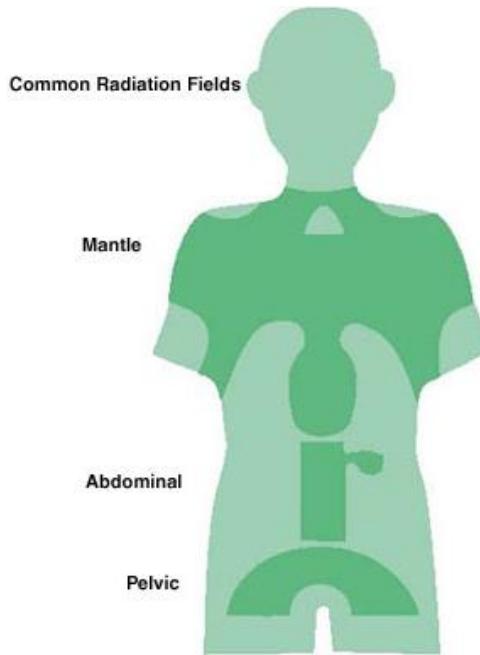


Amplified cells are radioresistant



Multiple copies (<10) of *c-MYC*

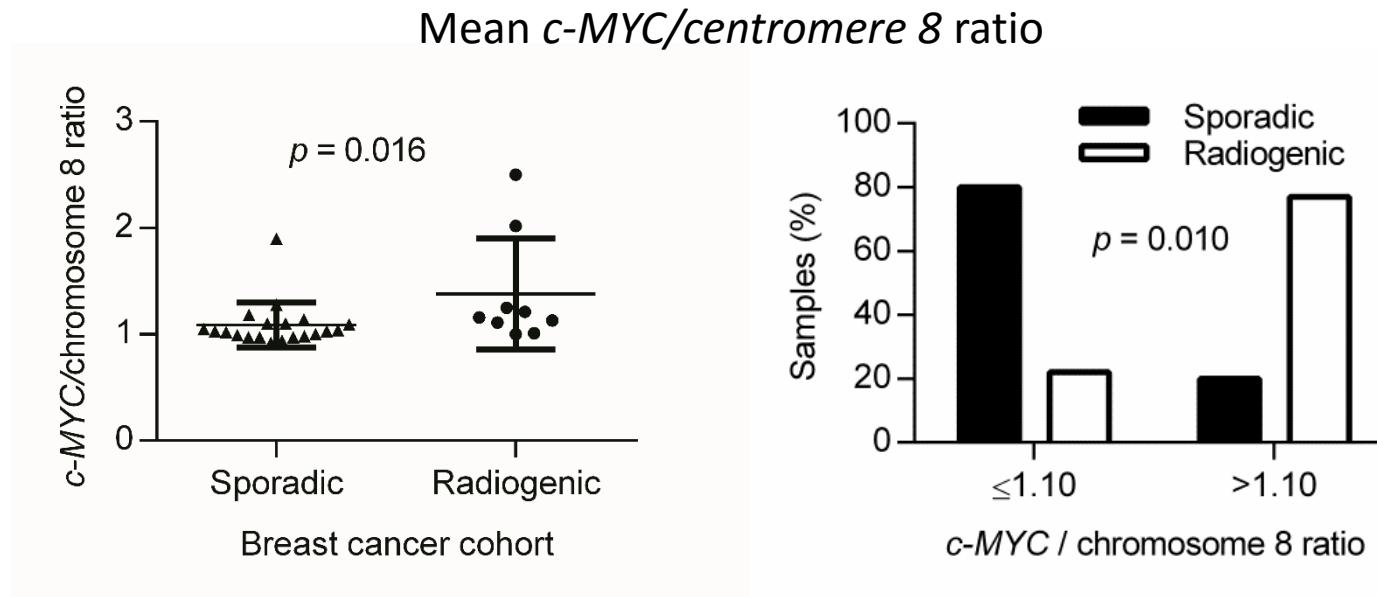
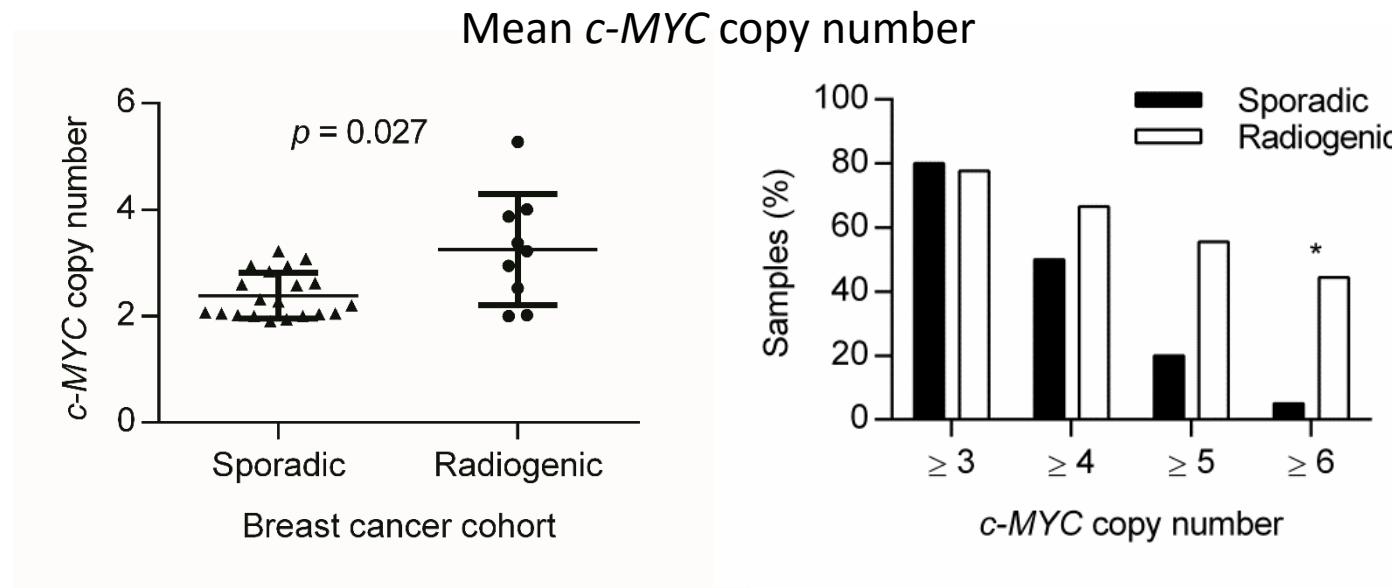
Breast cancer (BC) in HL survivors



- Breast cancer after radiotherapy for Hodgkin lymphoma
 - N=18
 - Mean age at BC diagnosis 37 years, range 28-47 years
 - Mean age at HL diagnosis 21 years, range 16-29
 - Mean latency between HL and BC 15 years, range 7-24 years
 - Mean cumulative RT dose 19.9 Gy, range 1.2-42.7

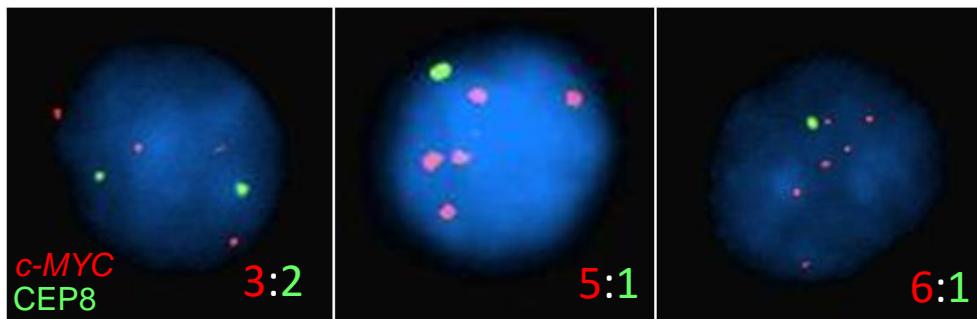
- Breast cancer without a radiation aetiology
 - N=33
 - Mean age at BC diagnosis 40 years, range 32-49 years.

c-MYC amplification in BC



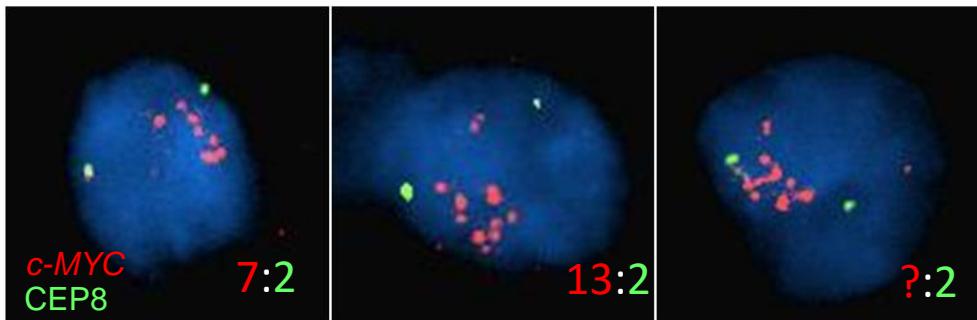
Heterogeneous c-MYC amplification

a SPO2



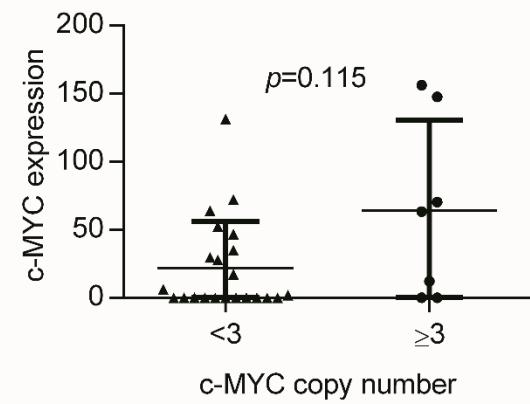
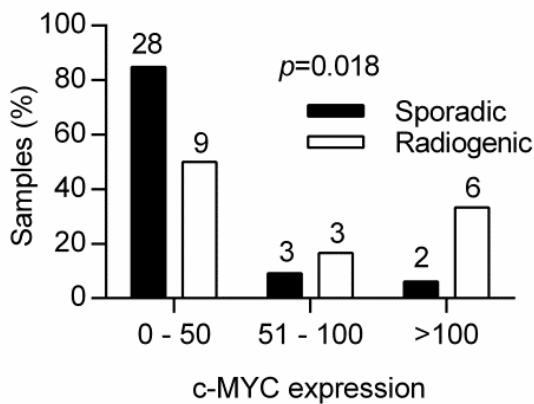
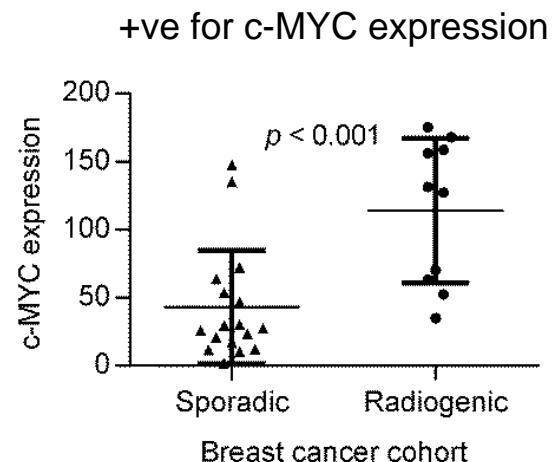
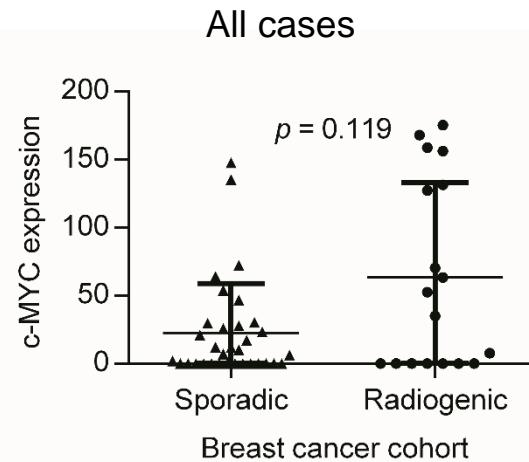
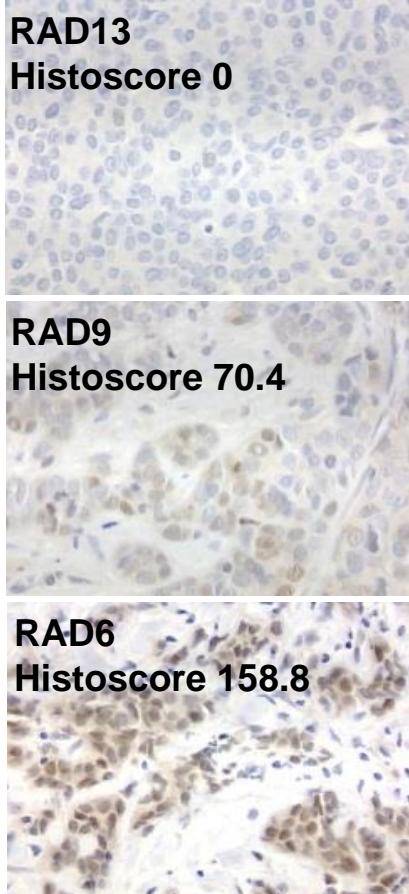
| | Copy number of probe ^A | | | | | | | | | | | | | |
|------|-----------------------------------|----|---|---|----|---|---|---|---|----|----|----|----|----|
| | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 | 13 | 14 |
| SPO2 | 5 | 53 | 8 | 6 | 21 | 5 | 2 | | | | | | | |
| | 38 | 62 | | | | | | | | | | | | |

d RAD10



| | Copy number of probe ^A | | | | | | | | | | | | | |
|-------|-----------------------------------|----|---|---|---|---|---|---|----|----|----|----|----|----|
| | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 | 13 | 14 |
| RAD10 | | 53 | | | 3 | 5 | 1 | 8 | 13 | 6 | 5 | 5 | | 1 |
| | 11 | 74 | 9 | 5 | 1 | | | | | | | | | |

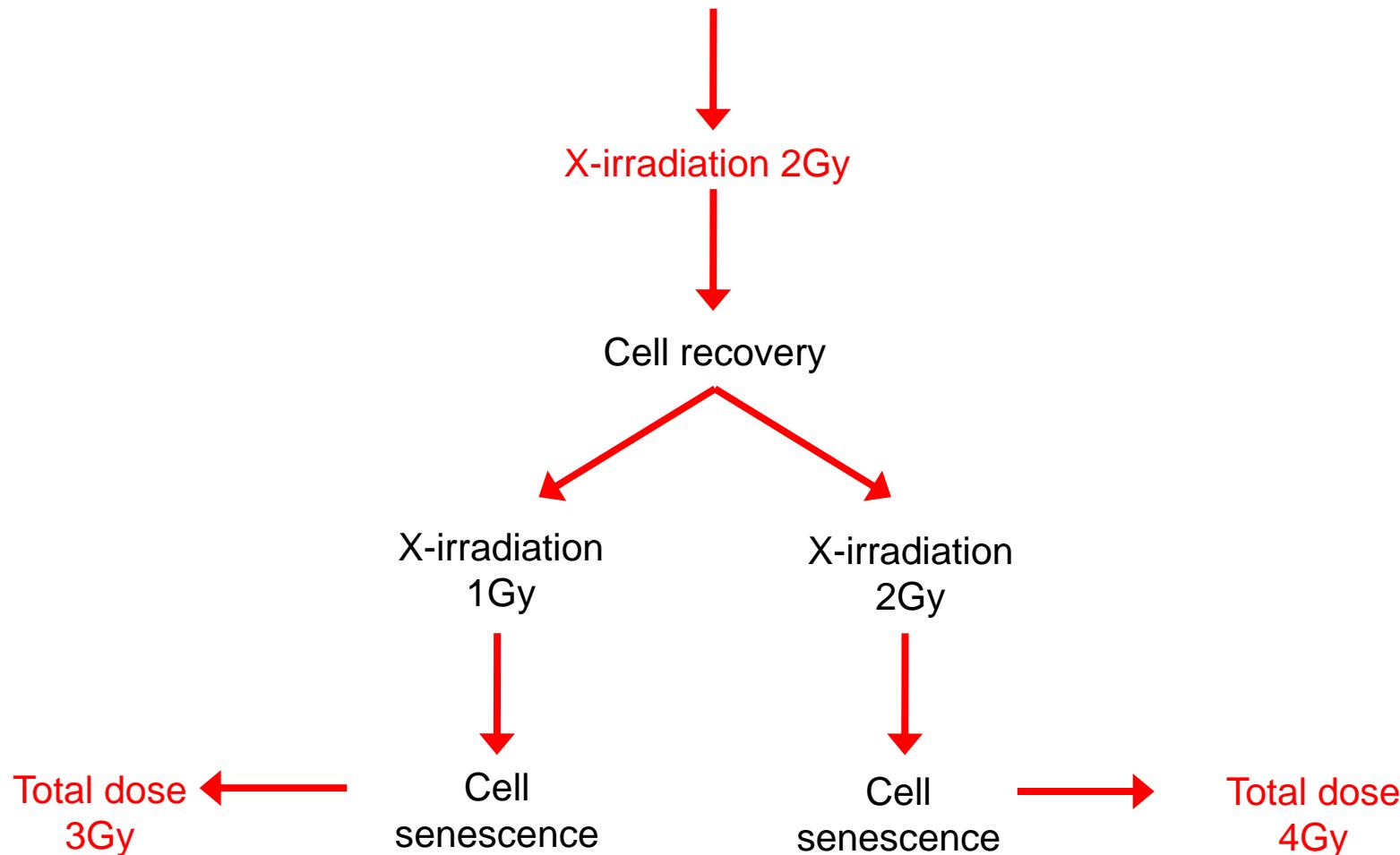
Radiogenic cancer has high c-MYC expression



Does radiation induce c-MYC amplification in non-immortalised primary cells?

Primary human mammary epithelial cells

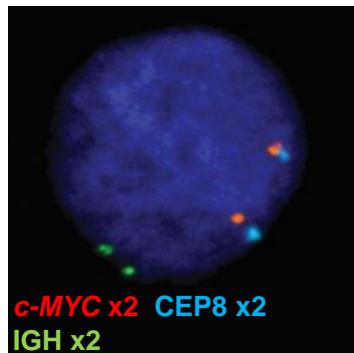
PRIMARY Human Mammary Epithelial Cells
(HuMEC)



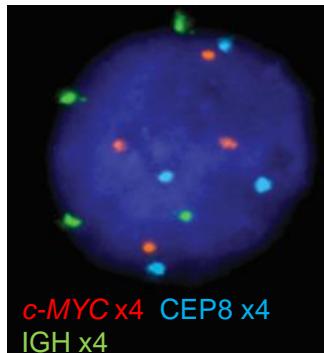
Primary human mammary epithelial cells

HuMEC FISH

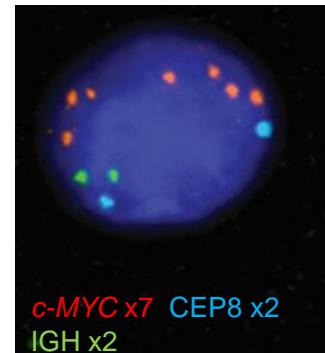
Non-amplified Diploid



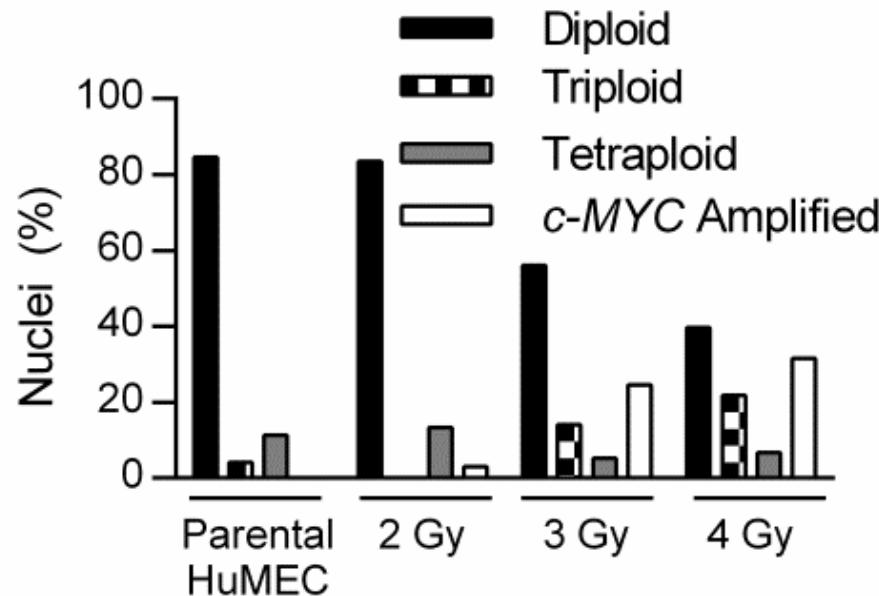
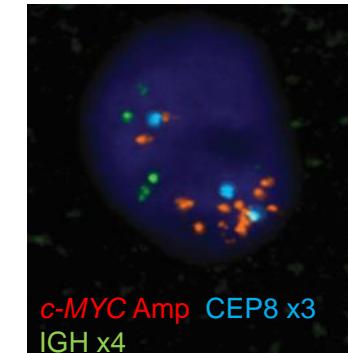
Tetraploid



Diploid Amplified



Polyplloid Amplified

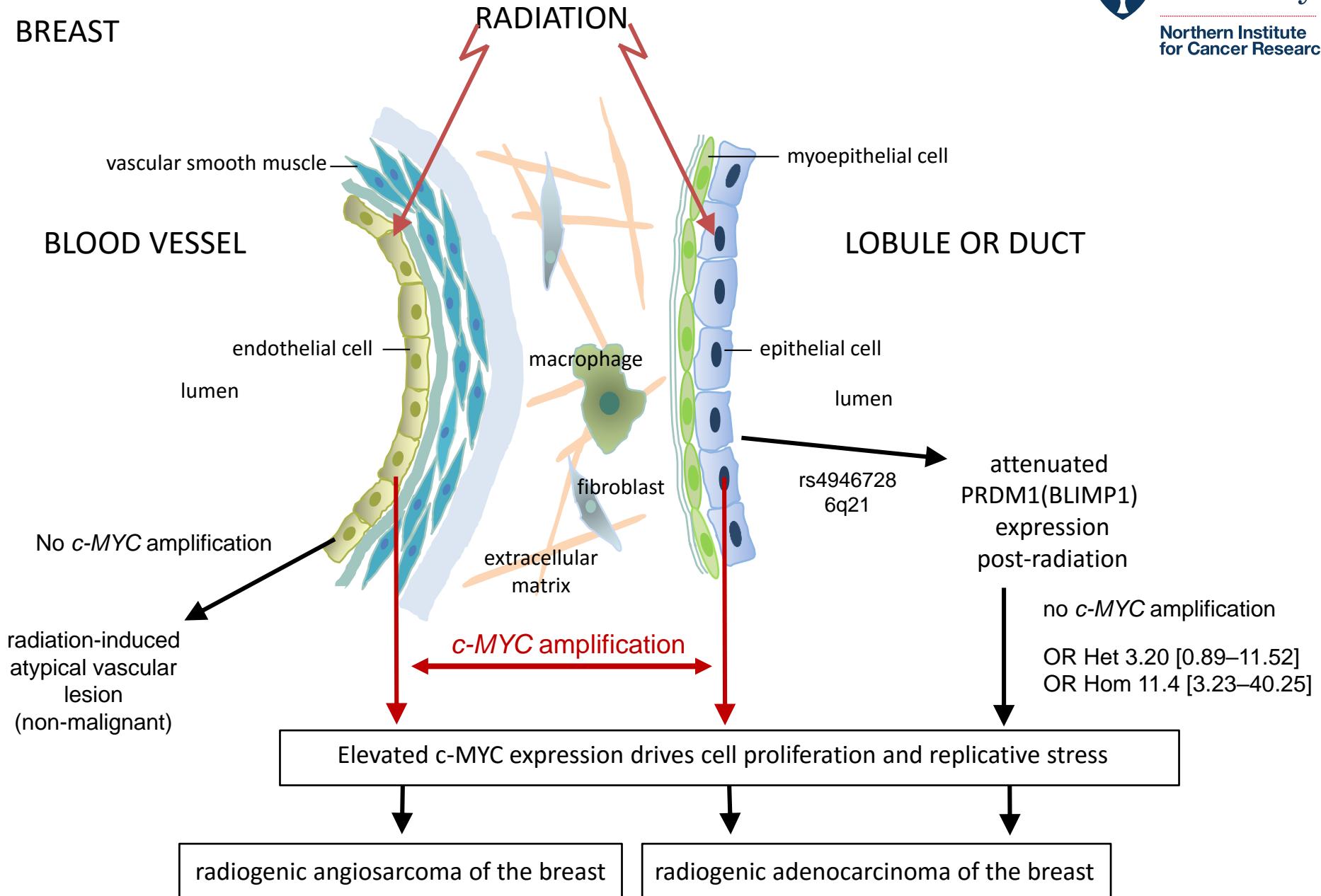


Model for radiogenic breast cancer

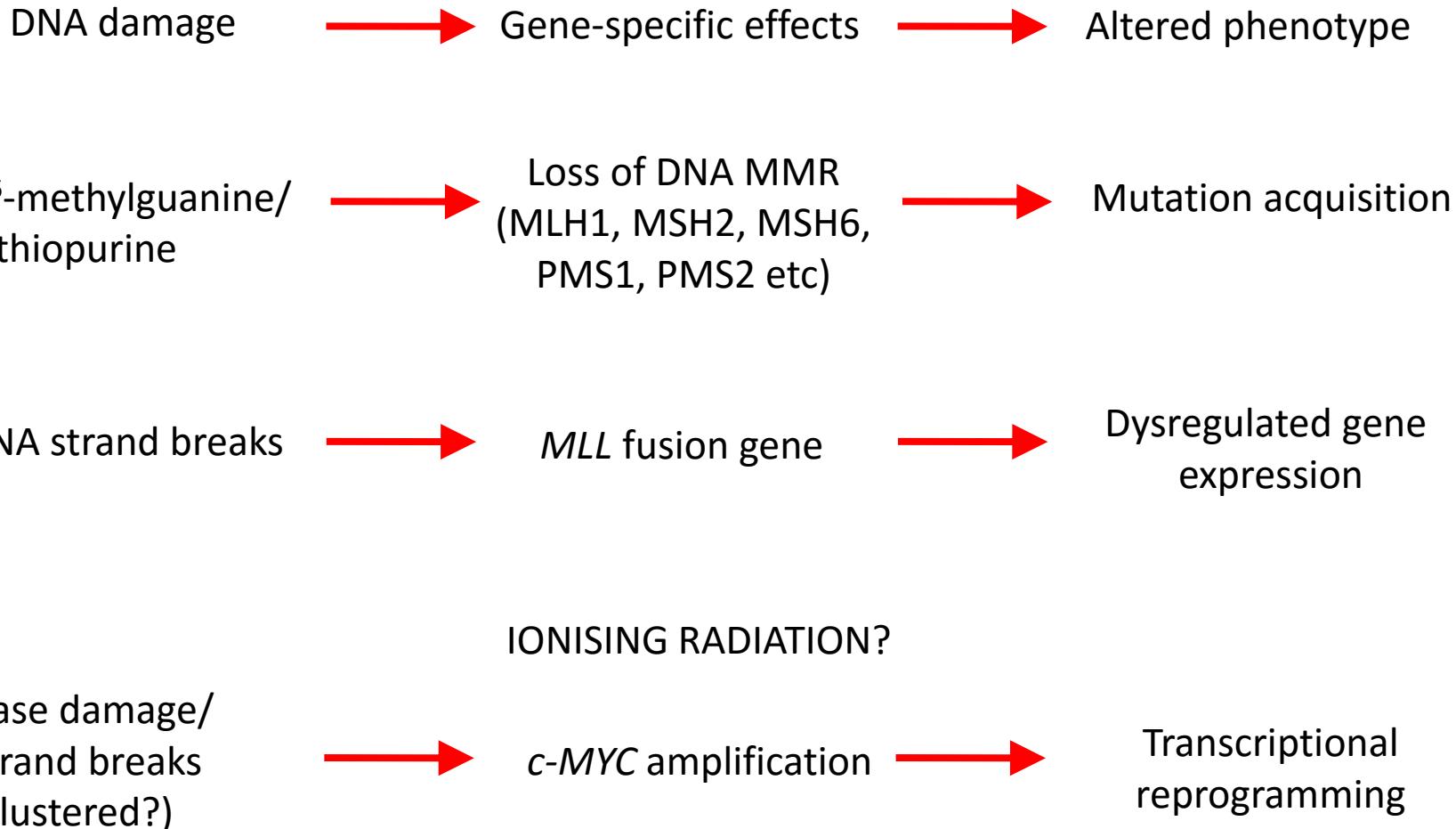
BREAST

BLOOD VESSEL

LOBULE OR DUCT

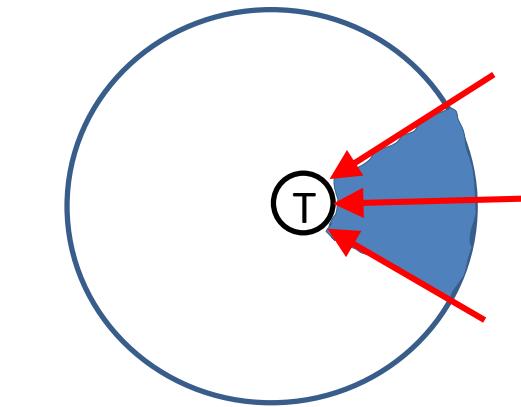


Molecular mechanisms of transformation

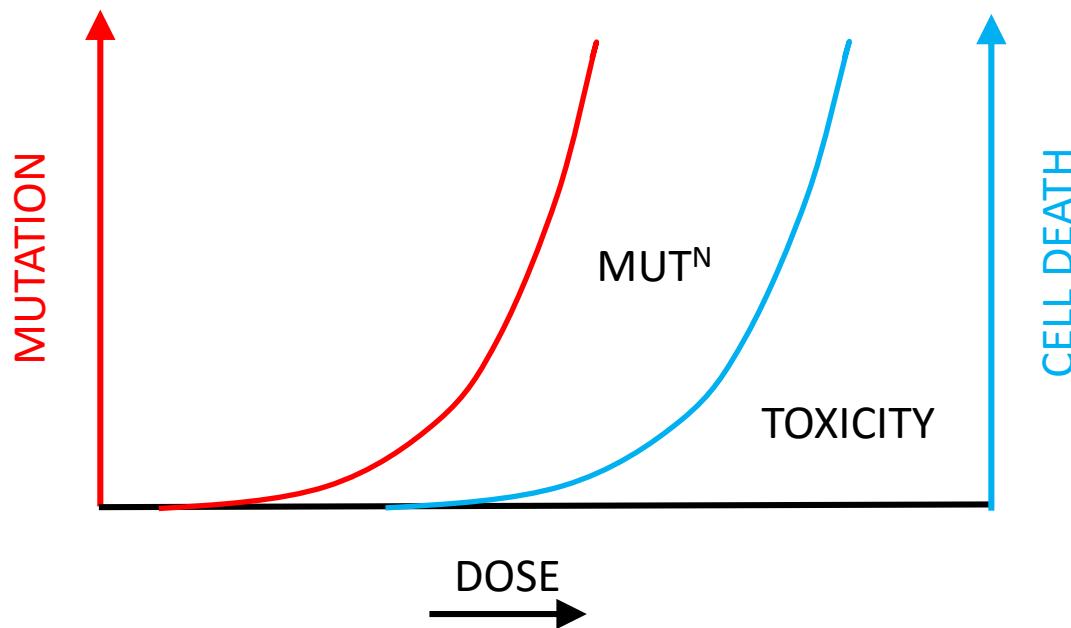


Risk modifiers?

Critical to minimise healthy tissue exposure

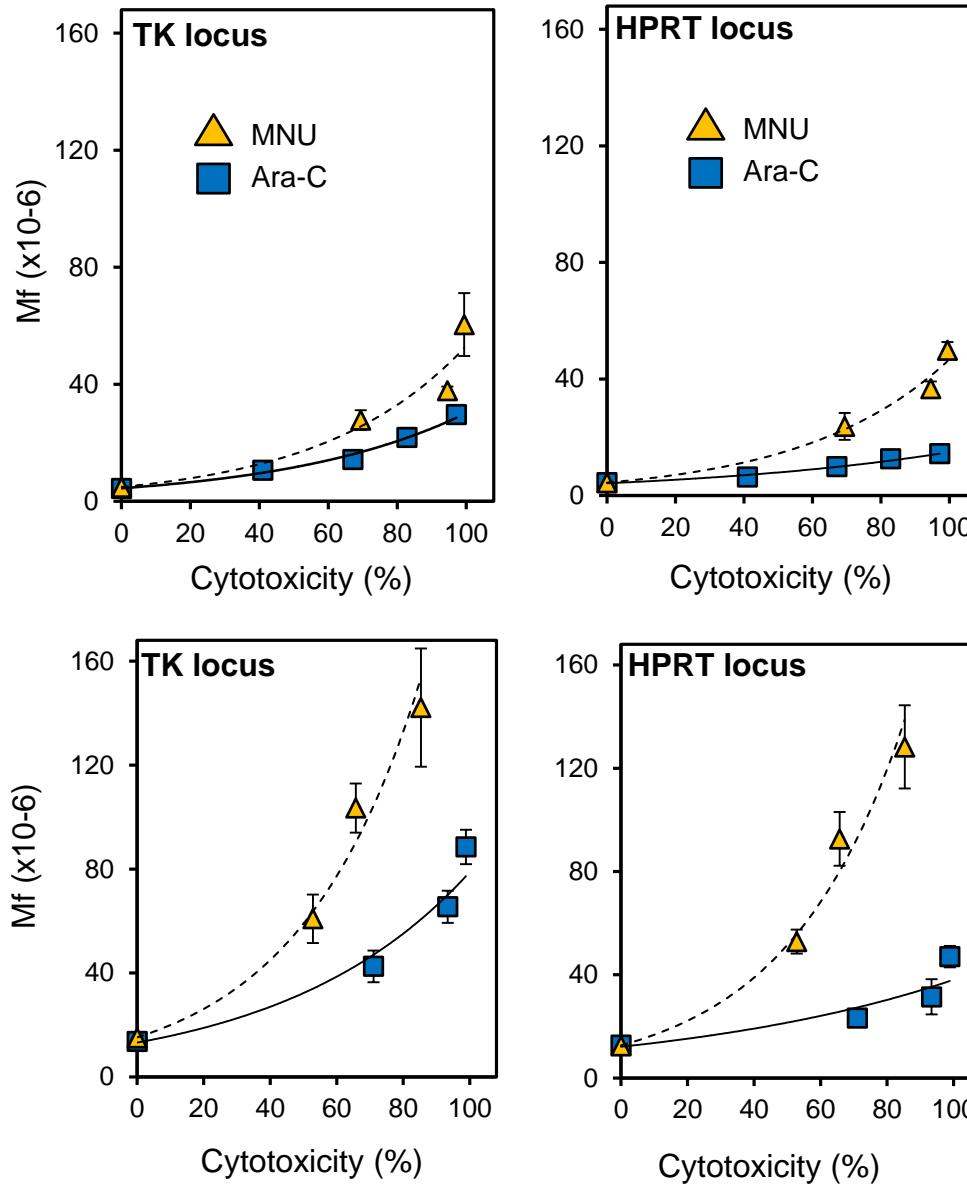
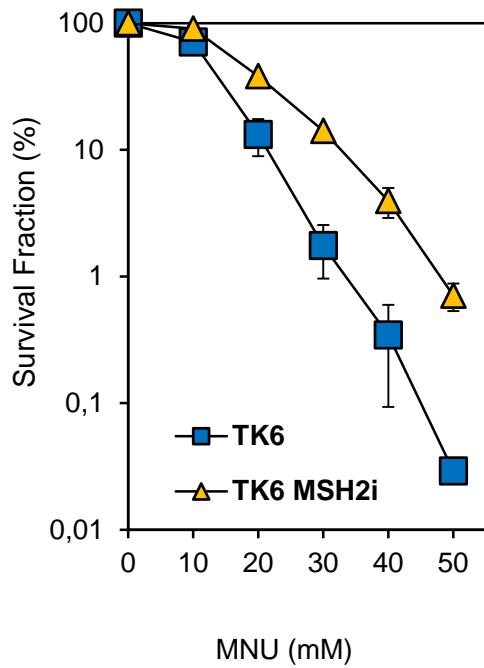


MUTATION CELL DEATH

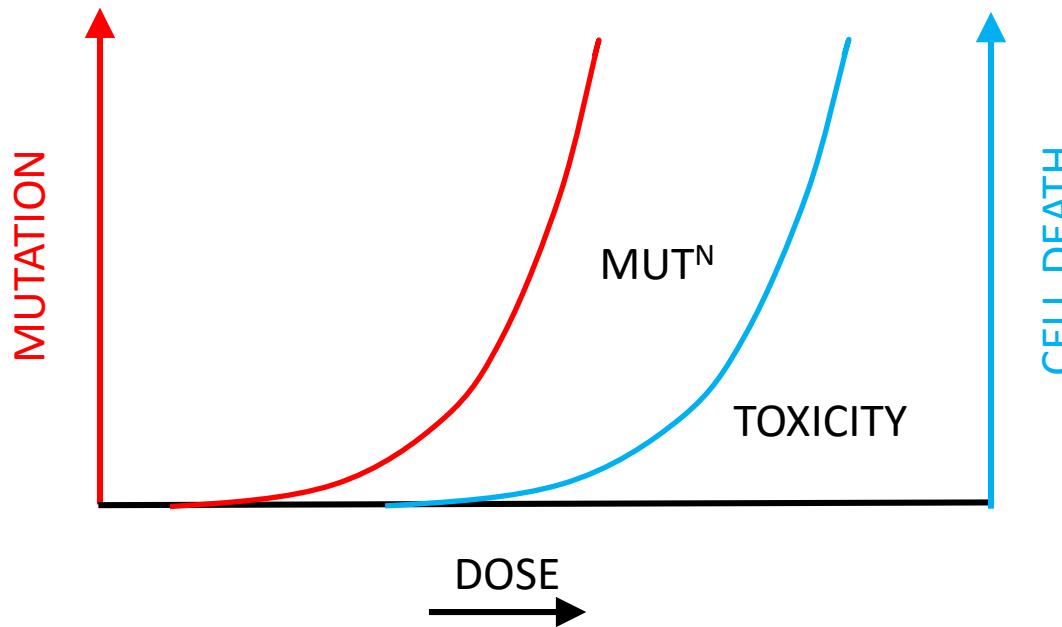


Identify modifiers that change the relationship between mutation and cell death

Chemotherapy-induced mutation



Loss of MSH2 increases the mutagenic window



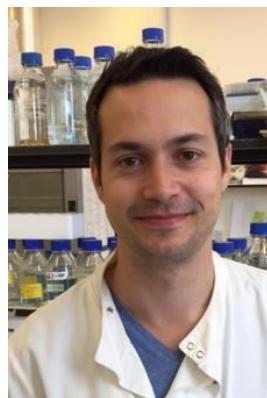
Identifying radiation risk modifiers?

- Secondary particles, bystander effects (likely mutagenic, are they also cytotoxic?)
- Proliferative index of the exposed healthy tissue (dividing cells fix more DNA to mutation)
- Volume of healthy tissue exposed (probability of SMN = mutations per cell X no. cells)
- Constitutional genetics (eg. *PRDM1* MAF 0.22; *RB1* MAF 10^{-5} , *TP53* rare)
- Acquired somatic alterations (DNA repair loss or apoptotic signalling dysfunction)
- Dose fractionation

Sarah Fordham



Mark Wade



Northern Institute
for Cancer Research

Newcastle
Sarah Fordham
Victoria Forster
Nicola Sunter
Helen Marr
Mohammed Nahari
Mark Wade
Felicity May

Chicago
Ken Onel

Indiana
Lois Travis



Newcastle
University

Bloodwise
Beating blood cancer since 1960

MRC | Medical
Research
Council

bright red
Fighting blood cancer
for a brighter future.

The Newcastle upon Tyne Hospitals 
NHS Foundation Trust